KINETIC RESOLUTION OF CHIRAL METALLOCENIC ALDEHYDES AND ALCOHOLS WITH LIVER ALCOHOL DEHYDROGENASE

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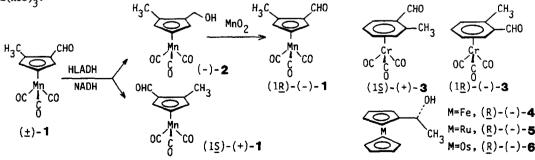
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Summary: Horse liver alcohol dehydrogenase-catalyzed oxidoreduction was useful to resolve racemic lformyl-2-methyl derivatives of tricarbonyl(cyclopentadienyl)manganese and (benzene)tricarbonylchromium and racemic l-hydroxyethylferrocene, ruthenocene and osmocene.

Enzyme-mediated asymmetric transformation is a useful method to prepare optically active organometallic compounds, as recently shown by us with horse liver alcohol dehydrogenase (HLADH) for ferrocene derivatives ^{la} and several other groups with hydrolases for 1-ferrocenylethanol, ^{lb,c} a dienoate-iron complex, ^{ld} or silicon-containing esters. ^{le} Microbial reductions of aromatic ketones and aldehydes complexed with $Cr(OO)_3$ have been also reported. ^{lf~h} Here we describe how HLADH is useful in the bioconversion of more various organometallic compounds.

(±)-Tricarbonyl ($?^{5}$ -1-formyl-2-methylcyclopentadienyl)manganese (1, 2 118 mg) was reduced with HLADH (120 U as assayed with EtOH) and NADH at pH 7.5 and 4°C. ³ When the TLC monitor showed that the spots for 1 and the product 2 had almost the same size at 2.5 hr, the compounds were extracted with EtOAc and isolated by silica gel column chromatography as optically active oils: 1 (36 mg, 31 % yield), (α)²⁶_D+101°(c=2.2, benzene); and 2 (41 mg, 35 % yield), (α)²⁵_D-8.7°(c=2.1, benzene).⁴ The latter was oxidized with MnO₂ to give the levorotatory aldehyde 1 in 78 % yield, (α)²³_D-104°(c=2.1, benzene). The absolute configuration of (+)-1 was determined to be (15, 2R) by its oxidation with Ag₂O to the known (15)-(+)-tricarbonyl($?^{5}$ -1-carboxy-2-methylcyclopentadienyl)manganese (76 % yield, mp 148~149°C, (α)²⁵_D+84°(c=0.50, EtOH); lit.⁵ mp 145~148°C, (α)⁶_D+83.3°(c=1.0, EtOH)). (±)-Tricarbonyl($?^{6}$ -2-methylbenzaldehyde)chromium (3)⁶ was also resolved by the HLADH-catalyzed

(±)-Tricarbonyl (η^{-2} -methylbenzaldehyde)chromium (3)° was also resolved by the HLADH-catalyzed reduction to (1<u>5</u>)-(+)-3. (28 % yield, mp 94~95°C, (α)_D²⁵+654°(c=0.20, CHCl₃); lit. mp 99~100°C,⁷ (α)_D²⁰+665°(c=0.3, CHCl₃)⁸) and (1<u>R</u>)-(-)-3. (mp 96~97°C, (α)_D²⁵-659°(c=0.20, CHCl₃)) via DMSO-Ac₂O oxidation of the enzymically produced 2-methylbenzyl alcohol complex (36 % yield, mp 97~98°C, (α)_D²⁵-12°(c=1.9, benzene)). The enantiomeric purity was almost 100 % e.e. for all aldehydes ((+)- and (-)-1 and 3), as evidenced by the single aldehyde proton signal in the PMR spectrum measured with Eu(hfc)₂.⁸



(t)-1-Hydroxyethylferrocene (4), ruthenocene (5) and osmocene (6) were enantioselectively oxidized with HLADH, NAD⁺ and FMN⁹ to give the ketones and levorotatory alcohols: (-)-4, 44 % yield, mp 76~77°C, $(\alpha)_D^{29}$ -29°(c=0.51, benzene) (lit.¹⁰ mp 72~73°C, $(\alpha)_D^{25}$ -30.5°(c=1.1, benzene)); (-)-5, 44 % yield, mp 77~78°C, $(\alpha)_D^{23}$ -21°(c=2.0, benzene) (lit.¹¹ $(\alpha)_D^{25}$ +20.7°(c=1.5, benzene)); and (-)-6, 33 % yield, mp 90~91°C, $(\alpha)_D^{23}$ -15°(c=2.0, benzene). HPLC analysis with a β-cyclodextrin-bonded column¹² indicated that the enantiomeric purity was 100 % e.e. for (-)-4 (with no detectable antipode peak) and 92 ± 1 % e.e. for (-)-5 and (-)-6.

The present enzymatic resolution method is very facile, as it needs neither derivatization such as the semioxamazone formation used for 3^7 nor temporary conversion to amine or carboxylic acid.^{11,14} Jaouen et al.^{1g} resolved an organometallic aldehyde resembling 3 by baker's yeast reduction, but the present HIADH method is more promising for high enantiomeric purity and experimental convenience.

References and Notes

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- 3. 1 was dissolved in the medium (60 ml of 0.1 M phosphate buffer, pH 7.5, containing 2 % EtOH and 2.5 % Tween 80) by sonication before addition of HLADH and NADH (85 µmol).
- 4. MS for (+)-1 m/z: 245.9717 (M⁺); calc. C₁₀H₇MnO₄=245.9725. MS for (-)-2 m/z: 247.9960 (M⁺); calc. C₁₀H₉MnO₄=247.9881. PMR (benzene-d₋) for (-)-2: The aldehyde proton signal (δ 9.18) of (+)-1 had disappeared and two coupled doublets were found at δ 3.73 and 3.83 (1H each, J=13 Hz, CHHOH).
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- 9. Typical reaction mixture: 180 ml of 0.1 M phosphate buffer (pH 8.3) containing 260 µmol NAD, 3.3 mmol FMN, 1.7 ml Tween 80 and 170 mg (±)-5. 800 U, 400 U and 400 U of the enzyme were added at 0, 30, and 53 hr, respectively. The mixture was stirred at 30 °C for 69 hr.
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